

MECHANISMS OF DISEASE

Chronic Lymphocytic Leukemia

Nicholas Chiorazzi, M.D., Kanti R. Rai, M.B., B.S., and Manlio Ferrarini, M.D.

From the Institute for Medical Research, North Shore–LIJ Health System (N.C., K.R.R.), and the Departments of Medicine, North Shore University Hospital, Manhasset, N.Y., and New York University School of Medicine, New York (N.C.); the Departments of Medicine, Long Island Jewish Medical Center, New Hyde Park, N.Y., and Albert Einstein School of Medicine, Bronx, N.Y. (K.R.R.); and the Division of Medical Oncology C, Istituto Nazionale per la Ricerca sul Cancro, and the Dipartimento di Oncologia Clinica e Sperimentale, Università di Genova — both in Genoa, Italy (M.F.). Address reprint requests to Dr. Chiorazzi at the Institute for Medical Research, North Shore–LIJ Health System, 350 Community Dr., Manhasset, NY 11030, or at nchizzi@nshs.edu.

N Engl J Med 2005;352:804-15.

Copyright © 2005 Massachusetts Medical Society.

When chronic lymphocytic leukemia (CLL) was last reviewed in the *Journal*,¹ it was considered a homogeneous disease of immature, immune-incompetent, minimally self-renewing B cells,² which accumulate relentlessly because of a faulty apoptotic mechanism.³ In the past decade, these views have been transformed by a wealth of new information about the leukemic cells. CLL is now viewed as two related entities, both originating from antigen-stimulated mature B lymphocytes, which either avoid death through the intercession of external signals or die by apoptosis, only to be replenished by proliferating precursor cells (Table 1).

normal b lymphocytes

B lymphocytes mature in the bone marrow (Fig. 1) and in the process rearrange immunoglobulin variable (V) gene segments to create the code for an immunoglobulin molecule that serves as the B-cell receptor for antigen. When an antigen of adequate affinity engages the receptor, the cell enters a germinal center in lymphoid follicles, where, as a centroblast, it rapidly divides and its V genes undergo somatic hypermutation (Fig. 2). This process introduces mutations in the rearranged V_HDJ_H and V_LJ_L gene segments that code for the binding site of the receptor. Through these mutations, the receptors of the descendant B cells, called centrocytes, acquire new properties. Cells with receptors that have enhanced antigen-binding affinity proliferate in the presence of the antigen, whereas centrocytes with receptors that no longer bind the antigen or do bind autoantigens are normally eliminated.⁴

This stimulation and selection pathway usually requires the help of T lymphocytes and occurs in germinal centers,⁴ the structure of which ensures the selection of antigen-avid B cells. However, the process can proceed without T cells⁵⁻⁸ and outside germinal centers, in the marginal zones around lymphoid follicles,⁹ most often in response to carbohydrates of encapsulated bacteria or viruses (Fig. 2). Both processes lead to the development of plasma cells or memory (antigen-experienced) B cells.

Concomitant with B-cell activation, the proteins on the surface of the B cell change. These modifications help activated B cells to interact with other cells and soluble mediators and thereby increase in number or mature into antibody-producing plasma cells. One surface molecule that supports B-cell interactions and differentiation is CD38.¹⁰ CD38 has adenosine diphosphate–ribose cyclase activity, and under certain circumstances augments signaling of B-cell receptors¹¹ and delivers signals that regulate the apoptosis of B cells.¹²

Signals received through B-cell surface receptors are transferred to the nucleus by a cascade of interacting molecules whose structures are temporarily modified during the process.^{13,14} These modifications frequently involve the attachment of phosphate groups to tyrosines of target proteins by specific enzymes. The enzymes involved in the initial phases of the signaling cascade include Syk and Lyn, members of the Src family

of protein tyrosine kinases.¹³ For T lymphocytes, the zeta-chain-associated protein 70 (ZAP-70) is a crucial player with similar activity.¹⁴

CLL and the biology of leukemic lymphocytes

The monoclonal population of B cells in CLL expresses CD19, CD5, and CD23 and has reduced levels of membrane IgM, IgD, and CD79b, a phenotype of mature, activated B lymphocytes.¹⁵⁻¹⁷ The pathological features of the lymph node are those of a small lymphocytic lymphoma.

Some patients with CLL survive for many years without therapy and eventually succumb to unrelated diseases, whereas others have a rapidly fatal disease despite aggressive therapy. Recognizing this heterogeneity, Rai and colleagues^{18,19} and Binet and colleagues²⁰ devised staging systems for use in assessing the extent of disease in an individual patient. These systems remain the cornerstones on which decisions regarding medical follow-up and treatment are built, but they fail to predict the course of the disease in patients in whom CLL is diagnosed in early stages.

Within the past decade, CLL has been shown to be a remarkably diverse disorder. Its heterogeneity reflects differences in the mutation status of V genes,²¹⁻²⁶ expression of CD38 and ZAP-70,²⁷⁻³⁵ and profiling of the expression of genes genome-wide.^{36,37} When cases of CLL were divided into categories on the basis of these differences, profoundly disparate clinical courses were revealed.^{25,26,28-30,38-40} Patients with clones having few or no V-gene mutations or with many CD38+ or ZAP-70+ B cells had an aggressive, usually fatal course, whereas patients with mutated clones or few CD38+ or ZAP-70+ B cells had an indolent course.

Mutations of V genes are detected by comparing DNA sequences of the genes in B cells with corresponding genes in the germ line. A sequence that differs from its germ-line counterpart by 2 percent or more is defined as mutated.²¹ According to this criterion, CLL cases are divisible into two groups: in the first, leukemic cells have rearranged V_H genes with 2 percent or more mutations ("mutated" CLL); in the second, there are few or no mutations (less than 2 percent; "unmutated" CLL).^{41,42} The presence of V-gene mutations and the presence of few CD38+ cells do not always correlate.^{26-28,43,44}

Table 1. A Comparison of Historical and Current Views of CLL.

Historical View	Current View*
CLL is a clinically heterogeneous disease with a homogeneous cellular origin.	CLL is a clinically heterogeneous disease originating from B lymphocytes that may differ in activation, maturation state, or cellular subgroup.
CLL is a disease derived from naive B lymphocytes.	CLL is a disease derived from antigen-experienced B lymphocytes that differ in the level of immunoglobulin V-gene mutations.
Leukemic-cell accumulation occurs because of an inherent apoptotic defect involving the entire mass of leukemic cells.	An inherent apoptotic defect involving the entire mass of leukemic cells is unlikely to exist initially. Leukemic-cell accumulation occurs because of survival signals delivered to a subgroup of leukemic cells from the external environment through a variety of receptors (e.g., B-cell receptors and chemokine and cytokine receptors) and their cell-bound and soluble ligands.
CLL is a disease of accumulation.	CLL is a disease of accumulation with a higher associated level of proliferation than was previously recognized.
Prognostic markers identify patients at various risk levels (low, intermediate, or high in the Rai staging categories and A, B, or C in the Binet categories) with an acknowledged heterogeneity in clinical outcomes among patients in the low- and intermediate-risk categories.	New molecular and protein markers identify patients within the low- and intermediate-risk categories who follow different clinical courses.
Therapy is based largely on clinical observations and trial-and-error methods.	New findings provide clues to discrete targets for developing hypothesis-driven and effective therapeutic agents.

* "Current view" refers to our understanding of the biology and derivation of CLL cells as it has evolved during the past 10 years.

ZAP-70 is an intracellular protein that promulgates activation signals delivered to T lymphocytes and natural killer cells by their surface receptors for antigen¹⁴ (Fig. 3). It is rarely present in normal B cells but has been found in B cells from patients with CLL.³⁶ When the expression of ZAP-70 is manipulated experimentally in B cells, it can facilitate signal transmission down the pathway initiated by antigen engagement with the B-cell receptor.⁴⁵ Gene-expression profiles indicate that unmutated CLL cells express more ZAP-70 mRNA than do mutated CLL cells.^{31-34,36} The analysis of DNA sequences to determine the status of immunoglobulin V-gene mutations is laborious and not performed routinely in clinical laboratories, whereas

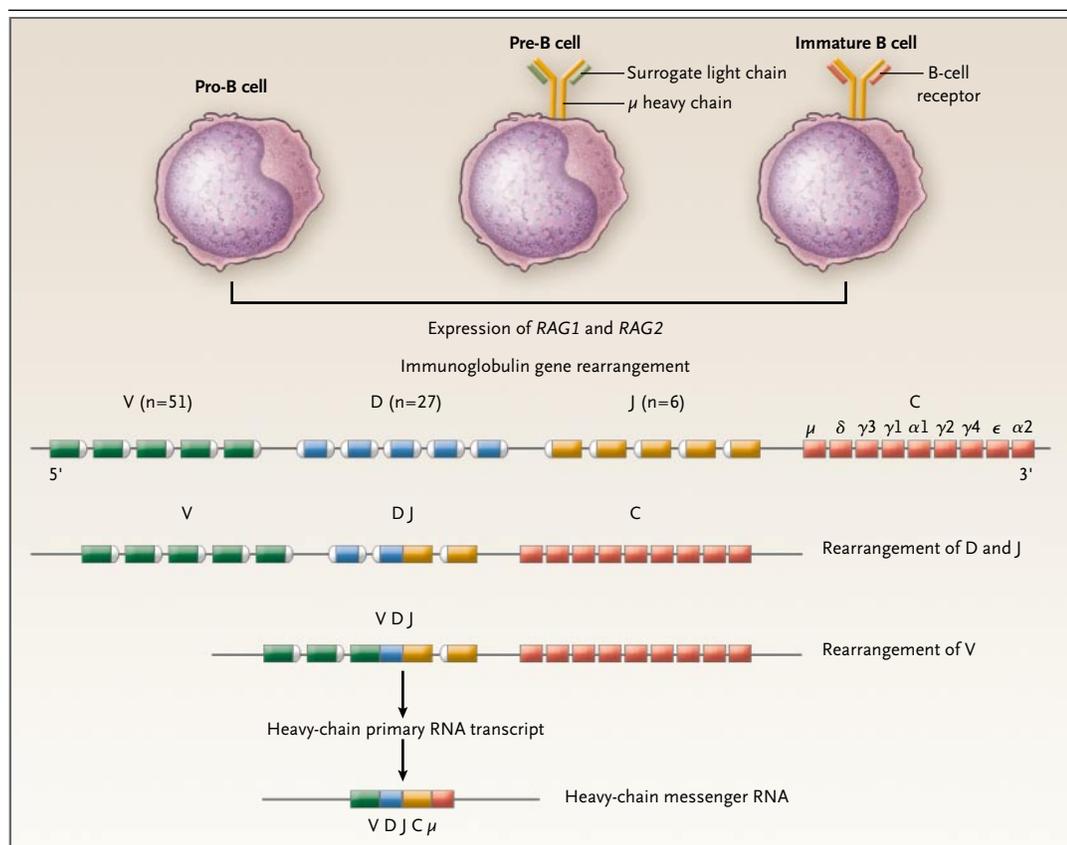


Figure 1. Normal Development of a B Lymphocyte.

Genetic units that will encode the variable (V) regions of the H and L (heavy and light) chains are rearranged in a step-wise fashion. Initially, 1 of the 27 D gene segments in the germ line links up with one of six J_H genes in pro-B cells. Next, 1 of the 51 V_H segments forms the DJ_H unit in pre-B cells. At this stage, the monomorphic pseudo-L chain (VpreB + λ5) is also synthesized, permitting the surface expression of the V_HDJ_H-μ + VpreB-λ5 complex (pre-B cell receptor). Similar recombination events occur at the L chain locus in pre-B cells, although they involve only two gene segments (V_L and J_L, not shown). The recombinase-activating genes *RAG1* and *RAG2* are essential for these molecular events. In addition, the enzyme terminal deoxytransferase may introduce additional nucleotides at the D-J_H and V_H-DJ_H junctions, thereby increasing diversity of the V regions of developing B cells. Each B cell uses a single set of V_HDJ_H and V_LJ_L rearrangements to create its antigen-binding site, thus maintaining the unique B-cell-receptor structure that identifies an individual B-lymphocyte clone. Each of these rearrangements must lead to the formation of recombinant genes encoding an intact immunoglobulin molecule to allow the cell to progress to the next stage of development.

testing for ZAP-70, when appropriately standardized, can more readily serve as a clinical test.^{31,33-35}

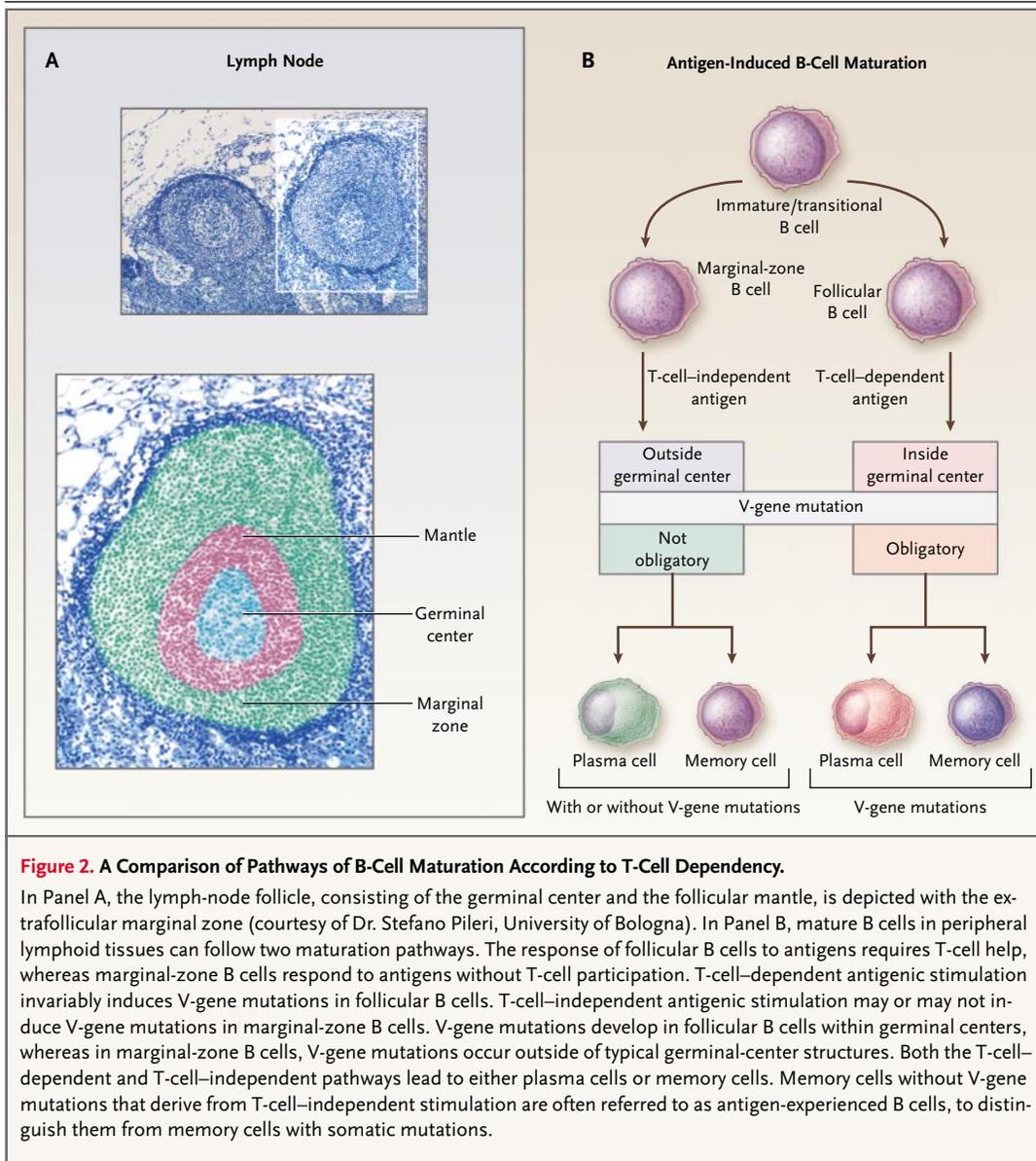
the biology of leukemic lymphocytes and the clinical course of CLL

INDUCING FACTORS IN CLL

Chromosomal translocations involving oncogenes frequently cause B-cell lymphomas.⁴⁶ CLL is a special case, however, because chromosomal translocations are rare, and no unifying mutations have been

identified. Yet the monoclonal nature of the B lymphocytes that proliferate in this disease imply that inducing lesions must exist in the progenitor clone.

Cytogenetic lesions are rare in the leukemic clone early in the course of the disease and therefore are not likely to be inducing factors. Nevertheless, some appear as the disease progresses. The most common is a deletion at 13q14.3, which occurs in more than 50 percent of cases over time.⁴⁷ This deleted region contains a nontranscribed gene⁴⁸ and two micro-RNA genes.⁴⁹ Micro-RNA is made normally by cells, including B lymphocytes,

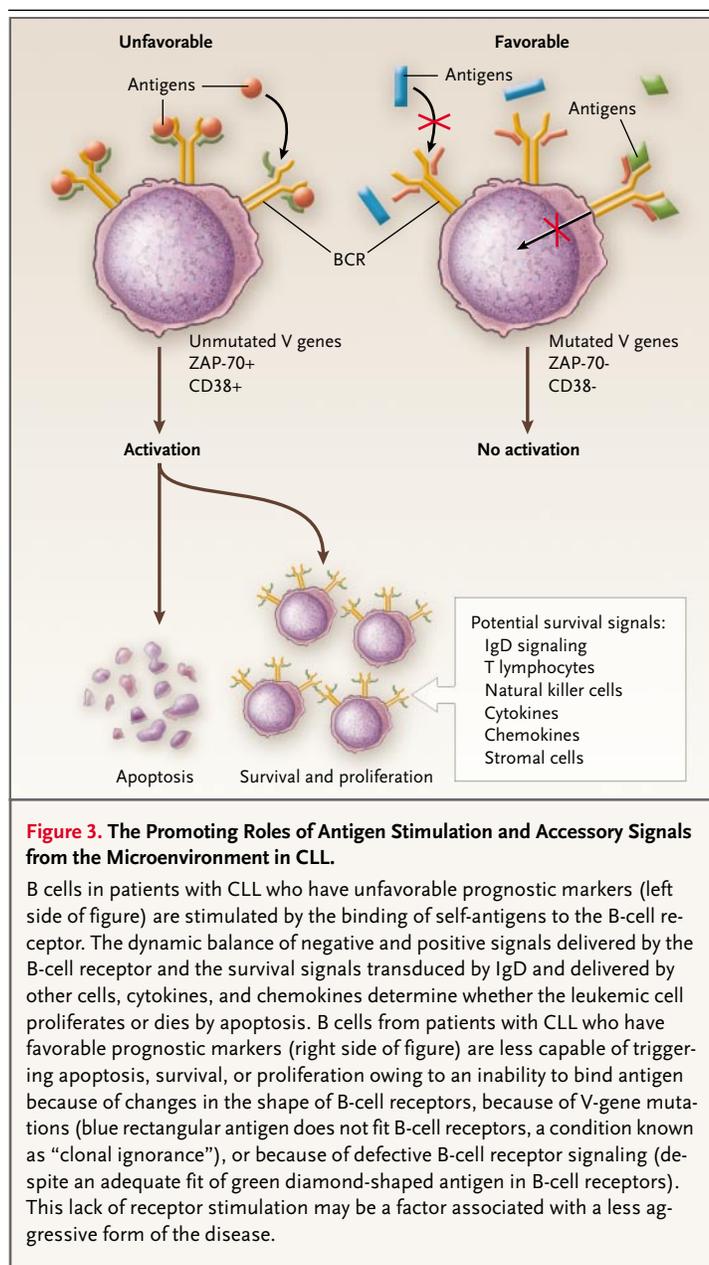


and regulate the functions of many genes,⁵⁰ some of which may have relevance to cancer in general and CLL in particular.⁵¹ Two micro-RNA genes located at 13q14 are deleted or down-regulated in most cases of CLL.⁵¹ The frequency of this deletion implies that it confers a selective advantage, possibly predisposing B-cell clones to undergo additional mutations.

The most ominous alterations are deletions at 11q22–23, 17p13, and 6q21.⁴⁷ Although the genes that are involved in these lesions are unknown, it is likely that p53 is included in the deletion at 17p13

and that the ataxia-telangiectasia mutated (ATM) gene is involved in the deletion at 11q22–23.^{47,52} Both genes regulate apoptosis and confer resistance to chemotherapy.⁵² These deletions are relatively frequent in unmutated CLL cases with a poor outcome.^{23,52}

Interestingly, TCL-1, located at 14q32.1 and involved in the pathogenesis of T-cell prolymphocytic leukemia,⁵³ is expressed in CLL.⁵⁴ In mice that were genetically manipulated to overexpress Tc1-1 in B cells, a leukemia or lymphoma of CD5+ B cells developed that was reminiscent of CLL.⁵⁵ Although



attractive candidates for an inducing factor, abnormalities of TCL-1 or its regulation have not been identified in patients with CLL.

PROMOTING FACTORS IN CLL

New evidence suggests that antigenic stimulation, along with interactions with accessory cells and cytokines, is a promoting factor that stimulates proliferation of CLL cells and allows them to avoid apoptosis. These effects may differ in distinct CLL

subgroups and thereby lead to the disparity in clinical outcomes among individual cases.

Inferring the Role of Antigenic Stimulation from B-Cell Receptors

The B-cell receptors of CLL cells from various patients are often structurally very similar, suggesting that the antigens these receptors bind are similar and relevant to the pathogenesis of CLL.^{41,42} The extent of similarity varies among groups of patients. In some cases, there are shared features in the portion of the antigen-binding site contributed by the H chain (V_H , D, and J_H genes).^{41,42,56} In these cases, each V_H gene exhibits special patterns of mutations and preferential combinations with particular D or J_H segments, which generate distinct features in the antigen-binding pocket.^{24,57} These V_HDJ_H rearrangements and characteristics of antigen-binding pockets differ from the much broader diversity found in B cells from normal persons.^{24,58,59}

In other groups of cases, the structural similarity of the receptors involves the entire antigen-binding site, coded by both the H and L chains (V_H , D, and J_H , and V_L and J_L genes). In these instances, the receptors from various patients are very similar or virtually identical.⁶⁰⁻⁶⁵ As many as 10 percent of all CLL cases fall into distinct categories branded by receptors with structurally similar antigen-binding pockets; most are of the unmutated, poor-outcome type.⁶³ These findings are very striking since, given the number of possible combinations of V-gene segments that can encode antigen-binding domains, one would not expect to find 2 cases of CLL with such structurally similar B-cell receptors in more than 1 million cases.

These cases suggest that a limited set of antigens promotes division of the leukemic cells, increasing the likelihood of dangerous DNA mutations. What are these promoting antigens? They are unknown, but it is possible that latent viruses or commensal bacteria repetitively activate particular B-cell clones through the B-cell receptor. CLL would result, directly or indirectly, from specific infections and would be perpetuated by them — in a manner similar to the gastric lymphomas that evolve in response to *Helicobacter pylori*.⁶⁶

Alternatively, environmental antigens or autoantigens could provoke clonal expansion. CLL cells frequently have polyreactive receptors, which bind multiple antigens, including autoantigens,⁶⁷⁻⁷⁰ allowing stimulation by both autoantigens and microbial antigens. This mechanism is plausible for

unmutated CLL and also for a few cases of mutated CLL, since many unmutated^{71,72} and some mutated⁷³ immunoglobulin V genes encode such polyreactive receptors; this immune-stimulation mechanism is in keeping with the view that the basis of CLL is autoimmunization.⁷⁴

As constitutive low-level signaling is delivered through the B-cell receptor in normal B lymphocytes, perhaps to maintain the memory response and the B-cell repertoire,^{75,76} antigen may not be necessary to continue clonal expansion—antigen-independent triggering might occur through the B-cell-receptor signaling pathway because of another genetic lesion.

Signal Transduction after Antigen Engagement

For antigenic stimulation to underlie clonal expansion, the B-cell receptor must propagate an efficient signal to the cell nucleus (Fig. 3). Leukemic cells from different CLL subgroups can differ in this capacity. Cross-linking B-cell receptor molecules with antibodies to IgM in vitro mimics the engagement of antigens with B-cell receptors and transmits signals to the cell nucleus in approximately 50 percent of cases of CLL.⁷⁷⁻⁸¹ This phenomenon seems to occur mainly in unmutated CLL,^{27,32,82} but more patients need to be studied for this to be confirmed. CLL cells that do not respond to stimulation from the B-cell receptor may be frozen at a stage at which even normal B lymphocytes would be unresponsive to antigen.⁴¹ Alternatively, these cells could be anergic, possibly because of previous antigenic experience.⁴² Finally, these CLL cells may have become incapable of responding to antigens because of changes in the structure of their B-cell receptors caused by somatic mutations or an inability of the cells to come into contact with relevant antigens in vivo.⁴¹ Considering the number of cells that make up the leukemic clone in many patients (10^{11} to 10^{12}), it is likely that only a fraction of the members of the clone could encounter the antigen, especially if they are restricted to discrete anatomical locations or compartments.

Other, not mutually exclusive, possibilities to explain the lack of B-cell-receptor signaling include reduced numbers of B-cell-receptor molecules,¹⁷ uncoupling of the B-cell receptor from accessory molecules necessary for effective signal transduction,⁸³⁻⁸⁵ and mutations in these accessory structures.⁸⁶ It is interesting to note that responsiveness to stimuli delivered through surface IgD is frequently maintained in these cases.^{82,87}

Consequences of Signal Transduction through the B-Cell Receptor

Once signal transduction is initiated by the B-cell receptor, B lymphocytes progress into the cell cycle or die. Cross-linking of surface IgM in CLL cells that can transduce a signal can cause⁸¹ or prevent⁸⁸ apoptosis (Fig. 3), whereas cross-linking surface IgD invariably prevents apoptosis.^{12,81} This difference is unexpected, because the two surface isotypes express the same clone-specific antigen-binding site and provide concordant signals in mature B cells. The final outcome of B-cell receptor signaling in an individual CLL cell, therefore, depends on the balance between signals mediated by the two molecules.

Signals from the Microenvironment

Signals that are delivered by direct cell contact or soluble factors, which may or may not occur concomitantly with B-cell-receptor engagement, probably propagate the growth of CLL cells (Fig. 3). Interactions with stromal cells⁸⁹ or nurse-like cells⁹⁰ or interactions between CD38 and its natural ligand CD31⁹¹ rescue CLL cells from apoptosis in vitro and probably do the same in vivo. Activated T cells or other cells expressing CD40 ligand also support the growth of CLL cells.⁹² Finally, cytokines such as interleukin-4 and vascular endothelial growth factor⁹³⁻⁹⁵ and chemokines such as SDF-1⁹⁶ (particularly in the presence of stromal cells) support the expansion of CLL clones.

These signals tip the balance between anti-apoptotic signals and proapoptotic signals in favor of cell survival. There is up-regulation of the anti-apoptotic genes *BCL2*, *survivin*, and *MCL1* in leukemic cells.^{92,97} Rescue from apoptosis and facilitation of cell growth may occur preferentially in lymph-node pseudofollicles and bone marrow clusters,^{89,98} evidenced by expression of the cycling cell marker Ki-67 by the leukemic cells in these sites.⁹² Because growth of the clone depends on a variety of interactions with the environment, variations in the requirements for these interactions on part of the leukemic cells may be responsible for changes in the clinical course.^{99,100}

Appearance and Evolution of New Genetic Mutations

The emergence of new, aggressive clonal variants, which can worsen the disease, requires proliferation of the leukemic clone. In vivo studies using radioactive and nonradioactive means suggest that CLL cells are more dynamic than is usually appreci-

ated.¹⁰¹⁻¹⁰³ CLL cells have surprisingly brisk birth rates, ranging from about 0.1 to more than 1.0 percent of the clone per day.¹⁰³ If the total clonal burden of a typical patient with CLL is approximately 10^{12} cells, these birth rates point to the daily production of some 10^9 to 10^{10} new leukemic cells.

These rates of cell division are sufficient to permit clonal variants to emerge. Indeed, there is an association between brisk birth rates of CLL cells and progressive disease.¹⁰³ The rate of birth of leukemic cells, therefore, may be more relevant clinically than is either the blood lymphocyte count or the physical examination, since the lymphocyte count reflects the proliferative capacity of the leukemic cells and their potential to promote new DNA lesions, whereas the sizes of the lymph nodes and spleen on physical examination reflect a balance between cell proliferation and cell death. These findings may explain why telomeres, which cap and protect the ends of chromosomes but shorten with each cell division, are smaller in cells from patients in CLL subgroups that have poor outcomes.^{104,105}

a unifying hypothesis
for the development, growth,
and evolution of cll

GROWTH AND EVOLUTION OF CLL CELLS

The above considerations suggest a plausible model on which to build future hypotheses and studies. Stimulatory and growth signals from the environment of CLL cells allow them to avoid apoptosis and proliferate. These signals are delivered by the B-cell receptor, receptors for cytokines or chemokines and other ligands, and direct contact with accessory and stromal cells. The major growth effects mediated by the B-cell receptor appear to occur in cases in which the receptor permits binding of autoantigens and maintains the capacity to transmit stimulatory signals to the cell nucleus (i.e., those with unmutated and, to a lesser degree, mutated CLL B-cell receptors).

This model excludes an intrinsic apoptotic defect in all members of the leukemic clone. Indeed, *in vitro* observations demonstrate the absence of lesions in the major apoptotic pathways.^{87,106} Whether continued cell division is facilitated by external signals or not, the level of B-cell turnover *in vivo* can suffice to promote the development and outgrowth of subclones with new genetic lesions and a growth advantage (Fig. 4).

DEVELOPMENT OF CLL FROM NORMAL B LYMPHOCYTES

Many normal B lymphocytes with unmutated V genes produce antibodies capable of binding multiple antigens (e.g., carbohydrates, nucleic acids, and phospholipids)¹⁰⁷ and of providing the first line of defense against microorganisms.¹⁰⁸ If one of these cells contained or developed a genetic abnormality that allows it to resist restraint on clonal size (e.g., an initial inducing lesion), then this cell would be primed for leukemic transformation (Fig. 4A). Foreign antigens and autoantigens then could be important stimuli for the development of CLL.¹⁰⁹ B cells with such unmutated polyreactive B-cell receptors could expand and convert to CLL cells with repetitive exposure to microbes and to autoantigens (Fig. 4B).¹¹⁰⁻¹¹³ A similar mechanism may underlie the origin of mutated CLL, because V-gene mutations can occur without T-cell help^{5,7,8} outside of germinal centers,⁹ and these mutations can sometimes favor autoreactivity.⁷³ Such expansion would stop if V-gene mutations altered the structure of the B-cell receptor in a way that caused loss of binding to the stimulatory antigen (i.e., the development of “clonal ignorance”) (Fig. 4C).

This hypothesis implies that such expansions should be detectable in healthy patients. Recent studies suggest that small numbers of clonal B cells with the characteristics of CLL cells exist in the blood of approximately 3.5 percent of disease-free persons^{114,115} and in an even higher proportion of first-degree relatives of patients with CLL.¹¹⁶ Although studies of the B-cell receptors of these B-lymphocyte expansions are limited, initial information suggests that they are monoclonal and use some of the genes that encode the B-cell receptors of CLL clones.

FROM WHICH SUBPOPULATION DO CLL CELLS DEVELOP?

Since CLL cells resemble activated B lymphocytes,¹¹⁷ their cellular origin cannot be deduced solely from phenotypic analyses, a fact that makes it difficult to draw a direct parallel with B1 cells described in mice.¹¹⁸ However, certain functional features may help delineate their origin. Normal adult B cells that produce autoantibodies and antibodies against bacterial or viral carbohydrates reside in the marginal zone. It is possible that marginal-zone B cells are the precursors of both unmutated and mutated CLL cells, because B-cell

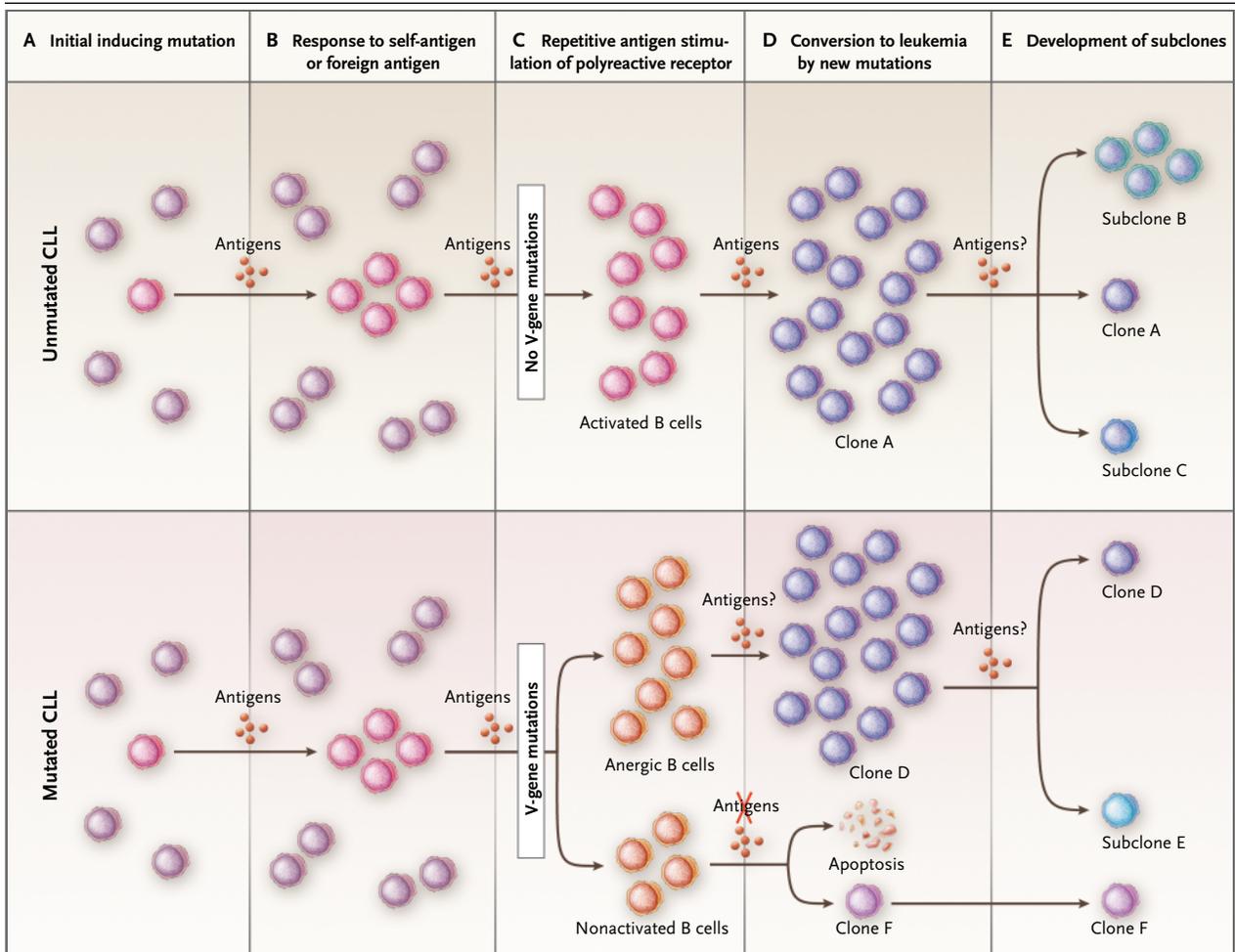


Figure 4. Model of the Development and Evolution of CLL Cells.

An initial inducing lesion occurs in a single B lymphocyte (pink cell in upper and lower portions of Panel A) among billions of distinct normal B cells. Interactions between antigens and B-cell receptors of adequate affinity induce clonal amplification (Panel B). The initial inducing lesion provides the marked cell with a growth advantage over other clones stimulated by the same or other antigens. In clones destined to become unmutated CLL cells, repetitive interactions between antigens and polyreactive B-cell receptors of the initially selected clone promote clonal growth, which persists because V-gene mutations do not occur (Panel C, top). In others, destined to become mutated CLL cells (Panel C, bottom), V-gene mutations develop that can abrogate the polyreactivity of the B-cell receptors and thereby alter their ability to bind the original antigen or autoantigen ("clonal ignorance"). Alternatively, these mutated cells become anergic owing to excessive B-cell–receptor stimulation because of the acquisition of more avid receptors. In both instances, the promoting effect of antigenic stimulation is neutralized. Additional DNA mutations cause the cells to cross the boundary from "normality" to "leukemia" (Panel D). Differences in the signals received through the B-cell receptors and other receptors determine the extent of clonal expansion. B cells that can no longer bind antigen may not cross this boundary. Continued cycling leads to other genetic changes (e.g., deletions at 13q, 11q, and 17p or duplication of chromosome 12) that determine the course of the disease (Panel E). These changes appear to occur more frequently in patients with unmutated CLL.

receptors of CLL cells are structurally similar to those of antibodies that react with autoantigens and carbohydrate components of infectious agents^{41,62,63} (Fig. 2B). Alternatively, mutated CLL cells could originate from B cells stimulated in a T-cell–dependent manner that have passed through a germinal center.

Other potential precursors are B1 cells, which share several features with marginal-zone B cells,¹¹⁸ as well as immature pre–B cells¹¹⁹ and transitional B cells¹²⁰ that can also express self-reactive receptors (Fig. 1). A few pre–B cells emerge from the bone marrow into the periphery, and transitional B cells routinely exit the marrow and traverse the

circulation to solid lymphoid tissues. A genetic abnormality could allow one of these cells to survive, thereby making it available for autoantigenic drive and leukemic transformation into unmutated CLL cells.

clinical implications

PROGNOSIS

The primary role of the Rai and Binet staging systems is to help clinicians decide when patients should be started on therapy. However, since these approaches do not predict the clinical course of a patient with precision, they are less helpful as long-term prognostic indicators. Therefore, physicians have postponed therapeutic decisions until the patients reach advanced Rai or Binet stages.

However, the molecular and cellular features we have discussed can distinguish patients with better or worse clinical courses, regardless of the Rai and Binet risk categories. Determination of V-gene mutation is not routinely available, but measurement of ZAP-70 is becoming widely available; it may be the most reliable indicator of prognosis.³⁵

Several points still require clarification and refinement. For example, can a single marker provide a sufficiently accurate and reliable prognostic assessment to permit early decisions about clinical management? Or should several markers be used to increase the degree of accuracy of prognosis? What are the clinically most useful cutoff points for the percentages of expression of CD38^{26,44} and ZAP-70³¹⁻³⁴ and the levels of immunoglobulin V-gene mutation²⁶ that most reliably define the clinical subgroups with various outcomes? Should the cases at the borderlines of these arbitrary cutoffs be handled differently?

MANAGEMENT

In the past, physicians told patients with CLL that a "watchful waiting" mode had to be adopted until the disease progressed, whereupon therapy would be initiated. In my opinion, this approach is especially disturbing, given that the novel prognostic markers indicate that some 50 percent of the patients assigned to watchful waiting have one or more features portending a poor outcome. Al-

though this approach is still being followed, it will probably change considerably when the new prognostic markers are more readily available to all clinicians. On the basis of such information, an early start of therapy may be justified in groups with a poor prognosis. However, before any guidelines can be proposed, the results of large, prospectively conducted clinical trials that test the use of early intervention in patients in poor-prognosis groups must become available. Only one such trial has been initiated, and the accrual of required numbers of patients and the analysis of those results will take several years.

NEW THERAPEUTIC APPROACHES

Since CLL cells must interact with the stroma in bone marrow or other peripheral lymphoid tissues to survive, these interactions need to be explored as targets of innovative therapies. Furthermore, specific inhibition of the B-cell receptor signaling pathway, in particular ZAP-70 or its signaling partners, may be an option. Targeting the actively proliferating cells that maintain the CLL clone by a cell-cycle-active agent could also be considered. Finally, since as many as 20 percent of patients with the worst prognostic markers have stereotypic antigen receptors, these common structures may be practical and valuable points of attack. When the antigens that engage these receptors are precisely defined, it may become possible to develop another arsenal of specific therapies.

Supported in part by grants from the National Cancer Institute (R01 CA 81554 and CA 87956) and a General Clinical Research Center Grant (M01 RR018535) from the National Center for Research Resources, the Associazione Italiana per la Ricerca sul Cancro, and the Ministero dell'Istruzione dell'Università e della Ricerca; the Peter J. Sharp Foundation, the Marks Family Foundation, the Jean Walton Fund for Lymphoma and Myeloma Research, the Joseph Eletto Leukemia Research Fund, the Tebil Foundation, the Horace W. Goldsmith Foundation, and the Chemotherapy Foundation. Drs. Rai and Chiorazzi are members of the NCI-sponsored Chronic Lymphocytic Leukemia Research Consortium (PO1 CA 081534).

Dr. Chiorazzi holds a patent on the use of CD38 as a prognostic indicator in chronic lymphocytic leukemia. Dr. Rai reports having received lecture fees from Genentech and Berlex Laboratories and grant support from Berlex. Dr. Ferrarini reports having received lecture fees from Schering AG.

We are indebted to the present and past members of the Laboratory of Experimental Immunology, Institute for Medical Research, North Shore-LIJ Health System, of the North Shore University Hospital, and of the Division of Medical Oncology C, Istituto Nazionale per la Ricerca sul Cancro, for the work that we have discussed in this article.

REFERENCES

1. Rozman C, Montserrat E. Chronic lymphocytic leukemia. *N Engl J Med* 1995;333:1052-7. [Erratum, *N Engl J Med* 1995;333:1515.]
2. Dameshek W. Chronic lymphocytic leukemia — an accumulative disease of immunologically incompetent lymphocytes. *Blood* 1967;29:Suppl:566-84.
3. Patel DV, Rai KR. Chronic lymphocytic leukemia. In: Hoffman R, Benz EJ, Shattil SJ, et al., eds. *Hematology, principles and practice*. 4th ed. Philadelphia: Elsevier Churchill Livingstone, 2005:1437-54.
4. Kelsø G. B cell diversification and differentiation in the periphery. *J Exp Med* 1994;180:5-6.
5. de Vinuesa CG, Cook MC, Ball J, et al. Germinal centers without T cells. *J Exp Med* 2000;191:485-94.
6. Monson NL, Foster SJ, Brezinschek HP, Brezinschek RJ, Dorner T, Lipsky PE. The role of CD40-CD40 ligand (CD154) interactions in immunoglobulin light chain repertoire generation and somatic mutation. *Clin Immunol* 2001;100:71-81.
7. Weller S, Faili A, Garcia C, et al. CD40-CD40L independent Ig gene hypermutation suggests a second B cell diversification pathway in humans. *Proc Natl Acad Sci U S A* 2001;98:1166-70.
8. Toellner KM, Jenkinson WE, Taylor DR, et al. Low-level hypermutation in T cell-independent germinal centers compared with high mutation rates associated with T cell-dependent germinal centers. *J Exp Med* 2002;195:383-9.
9. William J, Euler C, Christensen S, Shlomchik MJ. Evolution of autoantibody responses via somatic hypermutation outside of germinal centers. *Science* 2002;297:2066-70.
10. Deaglio S, Morra M, Mallone R, et al. Human CD38 (ADP-ribosyl cyclase) is a counter-receptor of CD31, an Ig superfamily member. *J Immunol* 1998;160:395-402.
11. Lund FE, Yu N, Kim KM, Reth M, Howard MC. Signaling through CD38 augments B cell antigen receptor (BCR) responses and is dependent on BCR expression. *J Immunol* 1996;157:1455-67.
12. Zupo S, Massara R, Dono M, et al. Apoptosis or plasma cell differentiation of CD38-positive B-chronic lymphocytic leukemia cells induced by cross-linking of surface IgM or IgD. *Blood* 2000;95:1199-206.
13. Niiron H, Clark EA. Regulation of B-cell fate by antigen-receptor signals. *Nat Rev Immunol* 2002;2:945-56.
14. Chan AC, Iwashima M, Turck CW, Weiss A. ZAP-70: a 70 kd protein-tyrosine kinase that associates with the TCR zeta chain. *Cell* 1992;71:649-62.
15. Matutes E, Owusu-Ankomah K, Morilla R, et al. The immunological profile of B-cell disorders and proposal of a scoring system for the diagnosis of CLL. *Leukemia* 1994;8:1640-5.
16. Moreau EJ, Matutes E, A'Hern RP, et al. Improvement of the chronic lymphocytic leukemia scoring system with the monoclonal antibody SN8 (CD79b). *Am J Clin Pathol* 1997;108:378-82.
17. TERNYNCK T, DIGHIERO G, FOLLEZOU J, BINET JL. Comparison of normal and CLL lymphocyte surface Ig determinants using peroxidase-labeled antibodies. I. Detection and quantitation of light chain determinants. *Blood* 1974;43:789-95.
18. Rai KR, Sawitsky A, Cronkite EP, Chanana AD, Levy RN, Pasternack BS. Clinical staging of chronic lymphocytic leukemia. *Blood* 1975;46:219-34.
19. Rai K. A critical analysis of staging in CLL. In: Gale RP, Rai KR, eds. *Chronic lymphocytic leukemia: recent progress, future direction*. New York: Alan R. Liss, 1987:253.
20. Binet JL, Auquier A, Dighiero G, et al. A new prognostic classification of chronic lymphocytic leukemia derived from a multivariate survival analysis. *Cancer* 1981;48:198-206.
21. Schroeder HW Jr, Dighiero G. The pathogenesis of chronic lymphocytic leukemia: analysis of the antibody repertoire. *Immunol Today* 1994;15:288-94.
22. Hashimoto S, Dono M, Wakai M, et al. Somatic diversification and selection of immunoglobulin heavy and light chain variable region genes in IgG+ CD5+ chronic lymphocytic leukemia B cells. *J Exp Med* 1995;181:1507-17.
23. Oscier DG, Thompson A, Zhu D, Stevenson FK. Differential rates of somatic hypermutation in V(H) genes among subsets of chronic lymphocytic leukemia defined by chromosomal abnormalities. *Blood* 1997;89:4153-60.
24. Fais F, Ghiotto F, Hashimoto S, et al. Chronic lymphocytic leukemia B cells express restricted sets of mutated and unmutated antigen receptors. *J Clin Invest* 1998;102:1515-25.
25. Hamblin TJ, Davis Z, Gardiner A, Oscier DG, Stevenson FK. Unmutated Ig V(H) genes are associated with a more aggressive form of chronic lymphocytic leukemia. *Blood* 1999;94:1848-54.
26. Krober A, Seiler T, Benner A, et al. V(H) mutation status, CD38 expression level, genomic aberrations, and survival in chronic lymphocytic leukemia. *Blood* 2002;100:1410-6.
27. Zupo S, Isnardi L, Megna M, et al. CD38 expression distinguishes two groups of B-cell chronic lymphocytic leukemias with different responses to anti-IgM antibodies and propensity to apoptosis. *Blood* 1996;88:1365-74.
28. Damle RN, Wasil T, Fais F, et al. Ig V gene mutation status and CD38 expression as novel prognostic indicators in chronic lymphocytic leukemia. *Blood* 1999;94:1840-7.
29. Ibrahim S, Keating M, Do KA, et al. CD38 expression as an important prognostic factor in B-cell chronic lymphocytic leukemia. *Blood* 2001;98:181-6.
30. Jelinek DF, Tschumper RC, Geyer SM, et al. Analysis of clonal B-cell CD38 and immunoglobulin variable region sequence status in relation to clinical outcome for B-chronic lymphocytic leukaemia. *Br J Haematol* 2001;115:854-61.
31. Crespo M, Bosch F, Villamor N, et al. ZAP-70 expression as a surrogate for immunoglobulin-variable-region mutations in chronic lymphocytic leukemia. *N Engl J Med* 2003;348:1764-75.
32. Chen L, Widhopf G, Huynh L, et al. Expression of ZAP-70 is associated with increased B-cell receptor signaling in chronic lymphocytic leukemia. *Blood* 2002;100:4609-14.
33. Wiestner A, Rosenwald A, Barry TS, et al. ZAP-70 expression identifies a chronic lymphocytic leukemia subtype with unmutated immunoglobulin genes, inferior clinical outcome, and distinct gene expression profile. *Blood* 2003;101:4944-51.
34. Orchard JA, Ibbotson RE, Davis Z, et al. ZAP-70 expression and prognosis in chronic lymphocytic leukaemia. *Lancet* 2004;363:105-11.
35. Rassenti LZ, Huynh L, Toy TL, et al. ZAP-70 compared with immunoglobulin heavy-chain gene mutation status as a predictor of disease progression in chronic lymphocytic leukemia. *N Engl J Med* 2004;351:893-901.
36. Rosenwald A, Alizadeh AA, Widhopf G, et al. Relation of gene expression phenotype to immunoglobulin mutation genotype in B cell chronic lymphocytic leukemia. *J Exp Med* 2001;194:1639-47.
37. Klein U, Tu Y, Stolovitzky GA, et al. Gene expression profiling of B cell chronic lymphocytic leukemia reveals a homogeneous phenotype related to memory B cells. *J Exp Med* 2001;194:1625-38.
38. Maloum K, Davi F, Merle-Beral H, et al. Expression of unmutated VH genes is a detrimental prognostic factor in chronic lymphocytic leukemia. *Blood* 2000;96:377-9.
39. Hamblin TJ, Orchard JA, Ibbotson RE, et al. CD38 expression and immunoglobulin variable region mutations are independent prognostic variables in chronic lymphocytic leukemia, but CD38 expression may vary during the course of the disease. *Blood* 2002;99:1023-9.
40. Oscier DG, Gardiner AC, Mould SJ, et al. Multivariate analysis of prognostic factors in CLL: clinical stage, IGVH gene mutational status, and loss or mutation of the p53 gene are independent prognostic factors. *Blood* 2002;100:1177-84.
41. Chiorazzi N, Ferrarini M. B cell chronic lymphocytic leukemia: lessons learned from studies of the B cell antigen receptor. *Annu Rev Immunol* 2003;21:841-94.
42. Stevenson FK, Caligaris-Cappio F.

- Chronic lymphocytic leukemia: revelations from the B-cell receptor. *Blood* 2004;103:4389-95.
43. Thunberg U, Johnson A, Roos G, et al. CD38 expression is a poor predictor for VH gene mutational status and prognosis in chronic lymphocytic leukemia. *Blood* 2001;97:1892-4.
44. Ghia P, Guida G, Stella S, et al. The pattern of CD38 expression defines a distinct subset of chronic lymphocytic leukemia (CLL) patients at risk of disease progression. *Blood* 2003;101:1262-9.
45. Kong GH, Bu JY, Kurosaki T, Shaw AS, Chan AC. Reconstitution of Syk function by the ZAP-70 protein tyrosine kinase. *Immunity* 1995;2:485-92.
46. Dalla-Favera R, Gaidano G. *Molecular biology of lymphomas*. 6th ed. Philadelphia: Lippincott, 2001.
47. Dohner H, Stilgenbauer S, Benner A, et al. Genomic aberrations and survival in chronic lymphocytic leukemia. *N Engl J Med* 2000;343:1910-6.
48. Migliazza A, Bosch F, Komatsu H, et al. Nucleotide sequence, transcription map, and mutation analysis of the 13q14 chromosomal region deleted in B-cell chronic lymphocytic leukemia. *Blood* 2001;97:2098-104.
49. Calin GA, Dumitru CD, Shimizu M, et al. Frequent deletions and down-regulation of micro-RNA genes miR15 and miR16 at 13q14 in chronic lymphocytic leukemia. *Proc Natl Acad Sci U S A* 2002;99:15524-9.
50. He L, Hannon GJ. MicroRNAs: small RNAs with a big role in gene regulation. *Nat Rev Genet* 2004;5:522-31.
51. Calin GA, Liu CG, Sevignani C, et al. MicroRNA profiling reveals distinct signatures in B cell chronic lymphocytic leukemias. *Proc Natl Acad Sci U S A* 2004;101:11755-60.
52. Stilgenbauer S, Lichter P, Dohner H. Genomic aberrations in B cell chronic lymphocytic leukemia. In: Cheson BD, ed. *Chronic lymphoid leukemias*. 2nd ed. New York: Marcel Dekker, 2001:353-76.
53. Virgilio L, Narducci MG, Isobe M, et al. Identification of the TCL1 gene involved in T-cell malignancies. *Proc Natl Acad Sci U S A* 1994;91:12530-4.
54. Narducci MG, Pescarmona E, Lazzeri C, et al. Regulation of TCL1 expression in B- and T-cell lymphomas and reactive lymphoid tissues. *Cancer Res* 2000;60:2095-100.
55. Bichi R, Shinton SA, Martin ES, et al. Human chronic lymphocytic leukemia modeled in mouse by targeted TCL1 expression. *Proc Natl Acad Sci U S A* 2002;99:6955-60.
56. Messmer BT, Albesiano E, Messmer D, Chiorazzi N. The pattern and distribution of immunoglobulin VH gene mutations in chronic lymphocytic leukemia B cells are consistent with the canonical somatic hypermutation process. *Blood* 2004;103:3490-5.
57. Johnson TA, Rassenti LZ, Kipps TJ. Ig VH1 genes expressed in B cell chronic lymphocytic leukemia exhibit distinctive molecular features. *J Immunol* 1997;158:235-46.
58. Widhopf GF II, Kipps TJ. Normal B cells express 51p1-encoded Ig heavy chains that are distinct from those expressed by chronic lymphocytic leukemia B cells. *J Immunol* 2001;166:95-102.
59. Potter KN, Orchard J, Critchley E, Mockridge CI, Jose A, Stevenson FK. Features of the overexpressed V1-69 genes in the unmutated subset of chronic lymphocytic leukemia are distinct from those in the healthy elderly repertoire. *Blood* 2003;101:3082-4.
60. Tobin G, Thunberg U, Johnson A, et al. Somatic mutated Ig V(H)3-21 genes characterize a new subset of chronic lymphocytic leukemia. *Blood* 2002;99:2262-4.
61. Tobin G, Thunberg U, Johnson A, et al. Chronic lymphocytic leukemias utilizing the VH3-21 gene display highly restricted V[lambda]2-14 gene use and homologous CDR3s: implicating recognition of a common antigen epitope. *Blood* 2003;101:4952-7.
62. Ghiotto F, Fais F, Valetto A, et al. Remarkably similar antigen receptors among a subset of patients with chronic lymphocytic leukemia. *J Clin Invest* 2004;113:1008-16.
63. Messmer BT, Albesiano E, Eftemov DG, et al. Multiple distinct sets of stereotyped antigen receptors indicate a key role for antigen in promoting chronic lymphocytic leukemia. *J Exp Med* 2004;200:519-25.
64. Tobin G, Thunberg U, Karlsson K, et al. Subsets with restricted immunoglobulin gene rearrangement features indicate a role for antigen selection in the development of chronic lymphocytic leukemia. *Blood* 2004;104:2879-85.
65. Widhopf GF II, Rassenti LZ, Toy TL, Gribben JG, Wierda WG, Kipps TJ. Chronic lymphocytic leukemia B cells of more than 1% of patients express virtually identical immunoglobulins. *Blood* 2004;104:2499-504.
66. Cavalli F, Isaacson PG, Gascoyne RD, Zucca E. MALT lymphomas. *Hematology (Am Soc Hematol Educ Program)* 2001:241-58.
67. Broker BM, Klajman A, Youinou P, et al. Chronic lymphocytic leukemic (CLL) cells secrete multispecific autoantibodies. *J Autoimmun* 1988;1:469-81.
68. Sthoeger ZM, Wakai M, Tse DB, et al. Production of autoantibodies by CD5-expressing B lymphocytes from patients with chronic lymphocytic leukemia. *J Exp Med* 1989;169:255-68.
69. Borche L, Lim A, Binet JL, Dighiero G. Evidence that chronic lymphocytic leukemia B lymphocytes are frequently committed to production of natural autoantibodies. *Blood* 1990;76:562-9.
70. Schwartz RS, Stollar BD. Heavy-chain directed B-cell maturation: continuous clonal selection beginning at the pre-B cell stage. *Immunol Today* 1994;15:27-32.
71. Diaw L, Magnac C, Pritsch O, Buckle M, Alzari PM, Dighiero G. Structural and affinity studies of IgM polyreactive natural autoantibodies. *J Immunol* 1997;158:968-76.
72. Dighiero G, Rose NR. Critical self-epitopes are key to the understanding of self-tolerance and autoimmunity. *Immunol Today* 1999;20:423-8.
73. Harindranath N, Goldfarb IS, Ikematsu H, et al. Complete sequence of the genes encoding the VH and VL regions of low- and high-affinity monoclonal IgM and IgA1 rheumatoid factors produced by CD5+ B cells from a rheumatoid arthritis patient. *Int Immunol* 1991;3:865-75.
74. Dameshek W, Schwartz RS. Leukemia and autoimmunization — some possible relationships. *Blood* 1959;14:1151-8.
75. Casola S, Otipoby KL, Alimzhanov M, et al. B cell receptor signal strength determines B cell fate. *Nat Immunol* 2004;5:317-27.
76. Monroe JG. Ligand-independent tonic signaling in B-cell receptor function. *Curr Opin Immunol* 2004;16:288-95.
77. Hivroz C, Grillo-Courvalin C, Brouet JC, Seligmann M. Heterogeneity of responsiveness of chronic lymphocytic leukemic B cells to B cell growth factor or interleukin 2. *Eur J Immunol* 1986;16:1001-4.
78. Karray S, Merle-Beral H, Vazquez A, Gerard JP, Debre P, Galanaud P. Functional heterogeneity of B-CLL lymphocytes: dissociated responsiveness to growth factors and distinct requirements for a first activation signal. *Blood* 1987;70:1105-10.
79. Michel F, Merle-Beral H, Legac E, Michel A, Debre P, Bismuth G. Defective calcium response in B-chronic lymphocytic leukemia cells: alteration of early protein tyrosine phosphorylation and of the mechanism responsible for cell calcium influx. *J Immunol* 1993;150:3624-33.
80. Lankester AC, van Schijndel GM, van der Schoot CE, van Oers MH, van Noesel CJ, van Lier RA. Antigen receptor nonresponsiveness in chronic lymphocytic leukemia B cells. *Blood* 1995;86:1090-7.
81. Zupo S, Cutrona G, Mangiola M, Ferrarini M. Role of surface IgM and IgD on survival of the cells from B-cell chronic lymphocytic leukemia. *Blood* 2002;99:2277-8.
82. Lanham S, Hamblin T, Oscier D, Ibbotson R, Stevenson F, Packham G. Differential signaling via surface IgM is associated with VH gene mutational status and CD38 expression in chronic lymphocytic leukemia. *Blood* 2003;101:1087-93.
83. Payelle-Brogard B, Magnac C, Alcover A, Roux P, Dighiero G. Defective assembly of the B-cell receptor chains accounts for its low expression in B-chronic lymphocytic leukaemia. *Br J Haematol* 2002;118:976-85.
84. Payelle-Brogard B, Magnac C, Oppezzo P, Dumas G, Dighiero G, Vuillier F. Retention and defective assembly of the B-cell receptor in the endoplasmic reticulum of chronic lymphocytic leukaemia B cells cannot be reverted upon CD40 ligand stimulation. *Leukemia* 2003;17:1196-9.

85. Alfaraano A, Indraccolo S, Circosta P, et al. An alternatively spliced form of CD79b gene may account for altered B-cell receptor expression in B-chronic lymphocytic leukemia. *Blood* 1999;93:2327-35.
86. Gordon MS, Kato RM, Lansigan F, Thompson AA, Wall R, Rawlings DJ. Aberrant B cell receptor signaling from B29 (Ig-beta, CD79b) gene mutations of chronic lymphocytic leukemia B cells. *Proc Natl Acad Sci U S A* 2000;97:5504-9.
87. Mangiola M, Cutrona G, Colombo M, et al. Mechanisms that control cell apoptosis or survival through surface Ig in B-CLL. *Leuk Lymphoma* 2003;44:Suppl:S4.
88. Bernal A, Pastore RD, Asgary Z, et al. Survival of leukemic B cells promoted by engagement of the antigen receptor. *Blood* 2001;98:3050-7.
89. Panayiotidis P, Jones D, Ganeshaguru K, Foroni L, Hoffbrand AV. Human bone marrow stromal cells prevent apoptosis and support the survival of chronic lymphocytic leukaemia cells in vitro. *Br J Haematol* 1996;92:97-103.
90. Tsukada N, Burger JA, Zvaifler NJ, Kipps TJ. Distinctive features of "nurselike" cells that differentiate in the context of chronic lymphocytic leukemia. *Blood* 2002;99:1030-7.
91. Deaglio S, Capobianco A, Bergui L, et al. CD38 is a signaling molecule in B-cell chronic lymphocytic leukemia cells. *Blood* 2003;102:2146-55.
92. Granziero L, Ghia P, Circosta P, et al. Survivin is expressed on CD40 stimulation and interfaces proliferation and apoptosis in B-cell chronic lymphocytic leukemia. *Blood* 2001;97:2777-83.
93. Orsini E, Foa R. Cytokines and regulatory molecules in the pathogenesis and clinical course of B-cell chronic lymphocytic leukemia. In: Cheson BD, ed. *Chronic lymphoid leukemias*. 2nd ed. New York: Marcel Dekker, 2001:127-60.
94. Chen H, Treweeke AT, West DC, et al. In vitro and in vivo production of vascular endothelial growth factor by chronic lymphocytic leukemia cells. *Blood* 2000;96:3181-7.
95. Lee YK, Bone ND, Stregre AK, Shanafelt TD, Jelinek DF, Kay NE. VEGF receptor phosphorylation status and apoptosis is modulated by a green tea component, epigallocatechin-3-gallate (EGCG), in B-cell chronic lymphocytic leukemia. *Blood* 2004;104:788-94.
96. Burger JA, Kipps TJ. Chemokine receptors and stromal cells in the homing and homeostasis of chronic lymphocytic leukemia B cells. *Leuk Lymphoma* 2002;43:461-6.
97. Pedersen IM, Kitada S, Leoni LM, et al. Protection of CLL B cells by a follicular dendritic cell line is dependent on induction of Mcl-1. *Blood* 2002;100:1795-801.
98. Ghia P, Caligaris-Cappio F. The indispensable role of microenvironment in the natural history of low-grade B-cell neoplasms. *Adv Cancer Res* 2000;79:157-73.
99. Lin K, Sherrington PD, Dennis M, Matrai Z, Cawley JC, Pettitt AR. Relationship between p53 dysfunction, CD38 expression, and IgV(H) mutation in chronic lymphocytic leukemia. *Blood* 2002;100:1404-9. [Erratum, *Blood* 2002;100:2291.]
100. Reed JC, Kitada S. Apoptosis dysregulation in chronic lymphocytic leukemia. In: Cheson BD, ed. *Chronic lymphoid leukemias*. 2nd ed. New York: Marcel Dekker, 2001:111-26.
101. Schiffer LM, Chanana AD, Cronkite EP, et al. Lymphocyte kinetics in chronic lymphocytic leukaemia (CLL) studied by ECIB. *Br J Haematol* 1969;17:408.
102. Theml H, Trepel G, Schick P, Kaboth W, Begemann H. Kinetics of lymphocytes in chronic lymphocytic leukemia: studies using continuous 3H-thymidine infusion in two patients. *Blood* 1973;42:623-36.
103. Messmer B, Messmer D, Allen SL, et al. In vivo measurements document the dynamic cellular kinetics of chronic lymphocytic leukemia B cells. *J Clin Invest* (in press).
104. Hultdin M, Rosenquist R, Thunberg U, et al. Association between telomere length and V(H) gene mutation status in chronic lymphocytic leukaemia: clinical and biological implications. *Br J Cancer* 2003;88:593-8.
105. Damle RN, Batiwalla FM, Ghiotto F, et al. Telomere length and telomerase activity delineate distinctive replicative features of the B-CLL subgroups defined by immunoglobulin V gene mutations. *Blood* 2004;103:375-82.
106. Chu P, Deforce D, Pedersen IM, et al. Latent sensitivity to Fas-mediated apoptosis after CD40 ligation may explain activity of CD154 gene therapy in chronic lymphocytic leukemia. *Proc Natl Acad Sci U S A* 2002;99:3854-9.
107. Boes M. Role of natural and immune IgM antibodies in immune responses. *Mol Immunol* 2000;37:1141-9.
108. Ochsenbein AF, Fehr T, Lutz C, et al. Control of early viral and bacterial distribution and disease by natural antibodies. *Science* 1999;286:2156-9.
109. Keating MJ, Chiorazzi N, Messmer B, et al. Biology and treatment of chronic lymphocytic leukemia. *Hematology (Am Soc Hematol Educ Program)* 2003:153-75.
110. Diaw L, Siwarski D, Coleman A, et al. Restricted immunoglobulin variable region (Ig V) gene expression accompanies secondary rearrangements of light chain Ig V genes in mouse plasmacytomas. *J Exp Med* 1999;190:1405-16.
111. Silverman GJ, Goldfien RD, Chen P, et al. Idiotypic and subgroup analysis of human monoclonal rheumatoid factors: implications for structural and genetic basis of autoantibodies in humans. *J Clin Invest* 1988;82:469-75.
112. Stevenson FK, Longhurst C, Chapman CJ, et al. Utilization of the VH4-21 gene segment by anti-DNA antibodies from patients with systemic lupus erythematosus. *J Autoimmun* 1993;6:809-25.
113. Pugh-Bernard AE, Silverman GJ, Cappione AJ, et al. Regulation of inherently autoreactive VH4-34 B cells in the maintenance of human B cell tolerance. *J Clin Invest* 2001;108:1061-70.
114. Rawstron AC, Green MJ, Kuzmicki A, et al. Monoclonal B lymphocytes with the characteristics of "indolent" chronic lymphocytic leukemia are present in 3.5% of adults with normal blood counts. *Blood* 2002;100:635-9.
115. Ghia P, Prato G, Scielzo C, et al. Monoclonal CD5+ and CD5- B-lymphocyte expansions are frequent in the peripheral blood of the elderly. *Blood* 2004;103:2337-42.
116. Rawstron AC, Yuille MR, Fuller J, et al. Inherited predisposition to CLL is detectable as subclinical monoclonal B-lymphocyte expansion. *Blood* 2002;100:2289-90.
117. Damle RN, Ghiotto F, Valetto A, et al. B-cell chronic lymphocytic leukemia cells express a surface membrane phenotype of activated, antigen-experienced B lymphocytes. *Blood* 2002;99:4087-93.
118. Martin F, Kearney JF. B1 cells: similarities and differences with other B cell subsets. *Curr Opin Immunol* 2001;13:195-201.
119. Meffre E, Schaefer A, Wardemann H, Wilson P, Davis E, Nussenzweig MC. Surrogate light chain expressing human peripheral B cells produce self-reactive antibodies. *J Exp Med* 2004;199:145-50.
120. Chung JB, Silverman M, Monroe JG. Transitional B cells: step by step towards immune competence. *Trends Immunol* 2003;24:343-9.

Copyright © 2005 Massachusetts Medical Society.